

10% FORMALDEHYDE

IVD *In vitro* diagnostic medical device



Fixative for use in histology INSTRUCTIONS FOR USE

REF Product code: F10-1L (1000 mL) F10-5L (5000 mL) F10-10L (10000 mL) F10-20L (20000 mL)

Introduction

An impeccable sample fixation is a prerequisite for a correct histological diagnosis. Tissue samples must be immersed in an optimally chosen fixative immediately after sampling, because a timely fixation will prevent autolysis, putrefaction and other unwanted cellular changes. Although there are hundreds of histological fixatives and at least tens of formaldehyde-based fixatives, neutral buffered formaldehyde solutions with a concentration range from 4% to 10% are the most commonly used fixatives, primarily because of their simple and universal application. Tissue fixation using a buffered formaldehyde solution results in forming cross-links, i. e. it forms methylene bridges between proteins, that is, it results in keeping tissue components in their *in vivo* relation. If fixated properly, the tissue sample can withstand additional histological tissue processing and staining. 10% formaldehyde solution is 25% formalin, which is the most commonly used fixative. Suitable for fixing larger tissue samples. It is a solution ready for use, colorless and with a characteristic odor, suitable for fixating larger tissue samples. It is a primary tissue fixative. Tissue samples can be fixated through a shorter period of time and with a lower volume ratio towards fixated tissue in comparison to 4% formaldehyde solution. Added methyl alcohol prevents formaldehyde polymerization. It is suitable for usage in all automated devices for tissue processing, as well as for manual histological techniques. It is conveniently packaged in 1 liter bottles, 5, 10 and 20 liter canisters.

Product description

- **10% FORMALDEHYDE** - 10% stabilized formaldehyde solution. Suitable for fixating larger tissue samples.

Fixating guidelines

If the tissue was not properly stored or stabilized during the fixation process, or if an unsuitable fixative was used, all the subsequent procedures in tissue processing and diagnostics will be of mediocre quality or useless. If the fixative is of inferior quality, pH value over its physiological bounds, or if the volume ratios between tissue and active substance in the fixative are not suitable, improper fixation can occur as well as tissue degradation and misdiagnosis. For that reason the fixative must be produced in accordance with the *in vitro* diagnostic devices norms and must bear the CE marking of conformity, and the processes of fixating, processing and staining must be carried out by a qualified person (histotechnician). In order to avoid mistakes during the procedure, a suitable fixative must be applied in accordance with the standard norms of histotechnology. If there is uncertainty regarding the choice of the fixative and possibility that the tissue would not be stored in a satisfactory manner, consultation with an experienced histotechnician is required.

Fixating instructions

- Always wear protective gloves while handling formaldehyde and fixated tissue samples. The rooms in which the formaldehyde is being used should be well ventilated by using an exhaust fan or a digester in order to remove toxic evaporation. Additional security information can be found in the Material Safety Data Sheet of this product.
 - Before the process a fixative should be chosen in accordance with the subsequent histological, histochemical or immunohistochemical diagnostic methods. If formaldehyde was chosen as an optimal fixative, the tissue sample should be immediately immersed in the solution container.
 - The sample should be fixated as soon as possible in order to prevent autolysis, putrefaction, and other changes. If it is not possible to put the sample in the fixative immediately, it is advised to maintain it moist and keep it in a cold place. The sample should not be bent or folded in the fixation container. Samples should be 3 to 6 mm in width for a proper fixation. All the samples should be clearly marked.
 - During the fixation the sample should be immersed in an adequate amount of fixative. An optimal ratio should be 20 to 40 parts of fixative to 1 part of tissue. That in particular applies to 4% Formaldehyde, while that ratio may be lower in case of 10% Formaldehyde. The fixative to tissue sample ratio should never be lower than 10 parts of fixative to 1 part of tissue.
 - If an entire organ is being fixated, the fixative should be injected into the organ or it can be cut into thin slices so that the solution can permeate the tissue thoroughly.
 - The fixative can also be poured into hollow organs, and before immersing into the fixative container they can be filled with gauze soaked with the fixative. Certain organs, such as the colon, can be opened and pinned on a board before immersing in the fixative. Encapsulated tissue should be processed by an expert in order for the fixation to be successful.
 - Fixation time can vary from a few hours to a few weeks. That depends on the type of tissue and sample thickness, fixation temperature, tissue and fixative volume ratio, as well as the concentration of formaldehyde in the fixative.
 - Selection of concentration of formaldehyde and fixation time must be determined in accordance with the norms of histotechnology and professional experience. In case of fixation of a larger tissue sample or an organ, fixation can last up to 24 hours or even more. The process can be shortened by fixating the sample in an incubator or a microwave oven.
- If the tissue has not been dimensioned for processing prior to fixation, after the fixation it should be processed down to thickness of 3-5 mm.

Preparing the sample and diagnostics

Use only appropriate instruments for collecting and preparing the samples. Process the samples with modern technology and mark them clearly. Follow the manufacturer's instructions for use. In order to avoid mistakes, the covering or mounting and staining procedure as well as diagnostics should only be conducted by authorized and qualified personnel. Use only microscope according to standards of the medical diagnostic laboratory. In order to avoid an erroneous result, a positive and negative check is advised before application.

Safety at work and environmental protection

Handle the product in accordance with safety at work and environmental protection guidelines. Used medical products and out of date products should be taken care of as a special waste in accordance with national guidelines. Chemicals used in this procedure could pose danger to human health. Tested tissue specimens are potentially infectious. Necessary safety measures for protecting human health should be taken in accordance with good laboratory practice. Act in accordance with signs and warnings notices printed on the product's label, as well as in BioGnost's material safety data sheet.

Storing, stability and expiry date

Keep Formaldehyde 10% in a tightly sealed original packaging at temperature of +15°C to +25°C. Do not keep in cold places, do not freeze and avoid exposing to direct sunlight. Date of manufacture and expiry date are printed on the product's label.

References

1. Carson, F. L., Hladik, C. (2009): *Histotechnology: A Self-Instructional Text*, 3rd ed., Chicago: ASCP Press
2. Cook, D. J. (2006): *Cellular Pathology*, 2nd ed., Banbury: Scion Publishing Ltd.
3. Kiernan, J.A. (2008): *Histological and Histochemical Methods, Theory and Practice*, 4th ed., Scion Publishing Ltd, Banbury.

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	Refer to the supplied documentation		Storage temperature range		Number of tests in package		Product code		European Conformity
	Refer to supplied instructions		Keep away from heat and sunlight		Valid until		Lot number		Manufacturer
	For <i>in vitro</i> diagnostic use only		Keep in dry place		Caution - fragile				



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